# Effect of PrP genotype on milk traits in Bovška sheep

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## **Abstract**

The aim of this study was to evaluate the effect of PrP genotype on milk traits in the Slovenian autochthonous Bovška sheep breed. The estimated population size is 3 700 animals, half of which are included in the selection programme. Genetic resistance for scrapie, a well known prion disease, is determined by the genotype at the PrP locus. In 2003, the Slovenian National Scrapie Programme following EU Regulations was introduced to control scrapie by selecting out the less resistant genotypes and until 2008, a total of 1875 ewes were genotyped. This study determined frequencies of PrP alleles in Bovška sheep, AHQ 17.32%, ARH 7.30%, ARQ 57.14%, ARR 17.12% and VRQ 1.12%, which were very similar to frequencies in other European breeds. The PrP genotype had a significant effect on milk, protein and lactose yield produced in lactation, but the productivity of the most resistant ARR/ARR homozygotes was similar to breed average and did not statistically differ from other genotypes. Heterozygotes AHQ/ARH, AHQ/ARQ and ARH/ARQ exhibited significantly higher productivity compared to respective homozygotes but not from the most resistant ARR/ARR genotype. It is expected therefore that increased frequency of the ARR allele or even its exclusivity will not directly influence the productivity of the breed. However, a careful management programme favouring resistant PrP genotypes should be implemented to reduce potential negative effects of productivity due to reduced selection potential or inbreeding.

**Keywords:** sheep, Bovška breed, PrP genotype, milk traits

# Zusammenfassung

# Einfluss der PrP Genotypen auf Milchmerkmale beim Bovška Schaf

Untersucht wurden der Einfluss der PrP Genotypen auf Milchmerkmale der bodenständigen Slowenischen Bovška Schafrasse. Der Rassenbestand wird auf 3 700 Tiere geschätzt, von denen die Hälfte der Population in das Zuchtprogramm einbezogen ist. Das Slowenische Scrapie National Programm startete 2003 auf der Basis der EU Regelung mit dem Ziel, die nicht resistenten Genotypen zu selektieren. Bis 2008 konnten in dieser Rasse 1 875 Mutterschafe genotypisiert werden. Die festgestellten Frequenzen betrugen: AHQ 17,32%, ARH 7,30%, ARQ 57,14%, ARR 17,12% sowie VRQ 1,12% und lagen damit ähnlich wie bei anderen europäischen Schafrassen. Der PrP Genotyp hatte einen signifikanten Einfluss auf den Milch-, Eiweiß- und Laktoseertrag. Die Leistungen der meist resistenten ARR/ARR homozygoten Genotypen ähnelten dem Rassendurchschnitt und unterschieden sich nicht signifikant von den anderen Genotypen. Die Heterozygoten AHQ/ARH, AHQ/ARQ und ARH/ARO zeigten signifikant höhere Leistungen im Vergleich zu

den Homozygoten außer zu ARR/ARR. Es ist zu erwarten, dass die Produktivität der Rasse durch eine höhere Frequenz oder ausschließlich ARR Allelen nicht beeinflusst wird. Für das Zuchtprogramm wird geschlussfolgert, dass bei einer Favorisierung der resistenten Allele vorsichtig zu verfahren ist und Auswirkungen hinsichtlich Produktion oder Inzucht zu beachten sind.

**Schlüsselwörter:** Schaf, Bovška Rasse, PrP Genotypen, Milchmerkmale

#### Introduction

Scrapie is a well known prion disease of sheep, which has occurred in many countries in the past, but not in the South, Central and Eastern European countries including Slovenia. However, recently it has spread into most European countries, including Slovenia. As early as 1962, it was speculated that scrapie was a genetic disorder (PERRY 1962). This hypothesis was later discounted because of observed infectivity of scrapie in sheep (DICKINSON et al. 1974) and transmission of the disease from sheep to goats (BROTHERSTON et al. 1968). However, DICKINSON et al. (1968) did note that some animals could be genetically resistant to scrapie. Nevertheless, it was not until 1995 that BELT et al. (1995) linked this phenomenon to the PrP locus. Five alleles responsible for different levels of susceptibility to scrapie were determined with the VRQ and ARR alleles being the most and least susceptible, respectively. Based on that observation, the European Commission has obligated (2003/100/EC) breeding organizations in the EU to implement selection programme to increase the frequency of ARR allele, and decrease the frequencies of less resistant alleles. However, homozygosity for the resistant ARR allele does not fully protect the animals against the infection and atypical cases of scrapie were also found in ARR homozygotes (LE DUR et al. 2005).

Intensive changing of allele frequencies at the PrP locus can potentially cause some negative consequences. Due to rapid selection for particular PrP genotypes, the genetic variability can be reduced. This may result in reduced selection potential or decreased production due to other negative effects such as inbreeding depression. These indirect negative consequences can be avoided with a well managed selection programme to prevent intensive reduction of less sensitive alleles like ARQ, ARH and AHQ. On the other hand, a direct influence of PrP genotypes on productivity is also possible. The PrP locus can influence productivity due to a linked quantitative trait locus or loci controlling some productivity trait. These linkages can be found in certain populations or breeds only. If the PrP genotype affects a quantitative trait, than the frequencies of PrP alleles are not random because selection changes allele frequencies at the PrP or linked quantitative trait locus.

The frequencies of PrP alleles differ from population to population. The ARQ is the most prevalent allele in most continental breeds ranging between 50.4% and 75.9% followed by the ARR allele ranging between18.2% and 42.5% (SALARIS *et al.* 2007, VAN KAAM *et al.* 2005, CASELLAS *et al.* 2006, BRANDSMA *et al.* 2004, BRANDSMA *et al.* 2005, DE MARCHI *et al.* 2005, ALVAREZ *et al.* 2006, WISNIEWSKA *et al.* 2006). However, in 7 of 16 breeds of Northern Germany, the ARQ allele was detected at a frequency below 41% while frequency of the ARR allele was higher than 50% in 5 of 16 breeds (DRÖGEMÜLLER *et al.* 2001).

Frequencies of the most sensitive VRQ allele are generally low in continental breeds ranging between 0.1% in Sardinian sheep (SALARIS *et al.* 2007) and up to 8.0% in the experimental Texel breed (BRANDSMA *et al.* 2005), with some outliers such as Cameroon sheep reaching a high 95% (DRÖGEMÜLLER *et al.* 2001). In comparison with continental breed, British populations differ widely in frequencies at the PrP locus. The ARR allele ranges between 31.1 and 76.5% (RODEN *et al.* 2006) and lower frequencies of ARQ compared to ARR allele were found in ten from eleven studied British breeds. An exception was the Scottish Blackface breed (ARQ between 14.3 and 58.9%), that also has low VRQ allele frequencies (0.3 to 6%). Similar frequencies were also found in New Zealand populations (BOSSERS *et al.* 1999), where the country is free of scrapie despite the fact that the populations originate from Great Britain.

The association studies of PrP alleles with production traits yielded contradictory results. In German black-headed breed, the non-ARR animals were found superior in muscle depth and daily gain (DE VRIES et al. 2003). On the other hand, PrP genotype did not significantly affect any milk trait in the Spanish Churra (ALVAREZ et al. 2006) and in the East Friesian dairy sheep (DE VRIES et al. 2005). However, the ARR homozygotes in Sardinian sheep produced lower milk yields (P < 0.05) than ARR heterozygotes or other genotypes (SALARIS et al. 2007). Thus, selection for the resistant ARR alleles could result in negative effects on milk productivity in Sardinian sheep. The PrP genotype did not show any association with reproduction traits in East Friesian dairy sheep (DE VRIES et al. 2005) or in Belclare ewes (SWEENEY et al. 2007). But the ARR/ARR and VRQ/- genotypes were associated with higher fertility in Texel breed (CASELLAS et al. 2007). Likewise, the ARH allele was superior compared to ARR and ARQ alleles in fertility and in weight gain in Ripollesa sheep (CASELLAS et al. 2007). The ARR/ARR genotype showed inferiority and the VRQ allele superiority in the Texel population for weight at 135 days of age (BRANDSMA et al. 2004) and on meat traits in the Ile de France, Prealpes du Sud, and Blanc du Massif Central breeds (VITEZICA et al. 2005) with no effects detected in the Texel experimental flock (BRANDSMA et al. 2006). The ARR locus did not influence fattening, meat guality and carcass traits of Thuringian breeds in Germany (GERNAND et al. 2005).

The literature reviewed above shows that quantitative traits are associated with PrP alleles in some but not all populations. Therefore, the consequences of selection for the ARR allele may differ from population to population. This implies that the effects of PrP genotypes on various productivity traits need to be studied and evaluated separately for each breed or population.

The Bovška sheep (Figure) is a Slovenian autochthonous dairy breed that originates from the Alpine north-western part of Slovenia and is also present at the Italian side of the border. In historical sources they refer to this breed as Krainer Steinschaf that probably originated from Zaupfelschaf (KAVAR *et al.* 2002). Narrow genetic distance between the Bovška sheep and another Slovenian alpine breed, Jezersko Solčava, suggested close relatedness (KAVAR *et al.* 2002). The wool of the Bovška breed is rough, mostly white, but can be also brown or black. Animals are of small stature with rams weighing between 45 and 50 and ewes between 35 and 40 kg. The head and ears have fine structure. These small animals are perfectly adapted for the poor conditions on alpine pasture and are efficient milk producers.



Bovška sheep breed
Bovška Schaf

### Material and methods

#### Animals and variables

Production data used in the present study originated from the Slovenian sheep breeding programme and have been collected since 1994. The estimated population of Bovška breed was 3 700 animals in year 2007 (EFABIS 2008). Selection for genetic resistance to scrapie started in the year 2006 and essentially all animals in the selection programme have been included in the genotyping scheme. Data used for this study were from 1 878 females genotyped from December 2004 up to April 10th, 2008 (34 in year 2004, 8 in year 2005, 705 in year 2006, 915 in year 2007 and 216 in year 2008). Only females with at least one finished lactation (in total 1568) were included in the statistical evaluation. Genotyped animals present almost half of the whole population. The genotypes were determined using ABI PRISM 7000 apparatus and the allelic discrimination assay.

The productivity of animals was measured according to the AT4 ICAR alternating method. The lactation length was calculated as the period between the lambing date and the date of drying out. Milk yield milked in lactation was calculated according to the following formula:

$$MY = \frac{\left(I_0 M_1 + \frac{I_1 (M_1 + M_2)}{2} + \dots + I_n M_n\right)}{1000} [kg]$$
 (1)

where MY is the milk yield in kg in lactation,  $I_0$  is an interval between the first day of lactation and the first milk recording day,  $I_1$  is an interval between the previous and the day of the next milk recording date,  $I_n$  is an interval between the last milk recording day and the day of drying out.  $M_1$  to  $M_n$  are the milk quantities, recorded at recording day one to the last recording day in g. The quantity of suckled milk in kg is estimated as five times the weight gain of a lamb or lambs from birth to weaning, where the birth weight of singles is estimated as 4 kg, and the birth weight of twins as 7 kg and the birth weight of triplets as 7.5 kg of live weight. Milk, produced in lactation is estimated as a sum of milk quantity, milked in lactation and milk quantity, suckled by lambs. The percentages of fat, proteins and lactose were analyzed. The percentage of dry matter was estimated as a sum of fat, proteins and lactose percentages. The quantity of fat, milk and lactose, produced in lactation was calculated from percentages and quantities of milk, produced in lactation.

#### Statistical methods

Statistical evaluation was done in three steps. In the first step, the frequencies of alleles and genotypes were calculated using SAS/STAT programme (SAS/STAT 2001) using procedure FREQ. The second step estimated average values of production data according to different genotypes by SAS/STAT programme, procedure MEANS. The third step evaluated analysis of variance. As the frequencies of genotypes VRQ/VRQ, AHQ/VRQ, ARR/VRQ, AHQ/AHQ and ARH/ARQ were very low, they were excluded from the analysis of variance, and so were flocks with less than 40 known PrP genotypes and animals with lactations shorter than 100 days.

The data set was evaluated with the following statistical model:

$$Y_{ijk} = \mu + G_i + F_j + b_1 \left( T_{ijk} - \overline{T} \right) + \sum_{l=1}^{3} b_{2l} \left( P_{ijk} - \overline{P} \right)^l + \sum_{m=1}^{3} b_{3m} \left( S_{ijk} - \overline{S} \right)^m + \sum_{n=1}^{2} b_{4n} \left( L_{ijk} - \overline{L} \right)^n + e_{ijk}$$
 (2)

where  $Y_{ijk}$  is an observation of the studied trait,  $\mu$  is an average of the statistical model,  $G_i$  is an effect of the i-th PrP genotype,  $F_j$  is an effect of the j-th flock (j-th farm),  $b_1(T_{ijk} - \overline{T})$  is a linear regression of year of lactation on a studied trait,  $\sum_{i=1}^{n} b_{i,i} (P_{ijk} - \overline{P})^i$  is a cubic regression effect of the day of lambing on every single studied trait and presents a seasonal effect,  $\sum_{i=1}^{n} b_{i,i} (\iota_{ijk} - \overline{\iota})^n$  is a quadratic regression of lactation length on studied traits and  $e_{ijk}$  is the residual of the observation ijk. The lambings are seasonal and therefore a day of lambing is simply the successive day in the year – January 1st had value 1, February 1st had value 32 etc.

The data were evaluated with SAS/STAT programme, procedure GLM that analyses the Least square means for genotypes. Differences between the expected values of heterozygotes, calculated as average values of two respective homozygotes and established values of heterozygotes (contrasts) were also calculated with the same programme.

### Results

#### **Frequencies**

Frequencies and percentages of PrP alleles as well as genotypes of Bovška breed are presented in Table 1. The most frequent allele in the Bovška breed is ARQ (57.14%). The proportion of the resistant ARR (17.12%) allele was similar to the proportion of the AHQ allele (17.32%). The proportion of the most sensitive VRQ allele was only 1.22%. The most sensitive genotype VRQ/VRQ was found only in one case (0.06%). Only 33 animals out of 1568 (2.11%) were heterozygotes possessing one VRQ allele. Homozygotes for the most resistant genotype ARR/ARR were more frequent (3.89%), however. More than a quarter of animals (25.98% or 406 animals) were heterozygotes with one ARR allele, ARR/ARQ, ARR/ARH and ARQ/ARQ.

Table 1
Frequencies (freq) and percentages (%) of PrP alleles and genotypes of Bovška breed
Frequenzen (freq) und Anteile (%) der PrP Allele und Genotypen der Bovška Schafe

					Genotype		
Allele			AHQ	ARH	ARQ	ARR	VRQ
AHQ	freq %	543 17.32	68 4.34	33 2.10	289 18.43	75 4.78	10 0.64
ARH	freq %	229 7.30		23 1.47	112 7.14	35 2.23	3 0.19
ARQ	freq %	1 792 57.14			542 34.57	296 18.88	11 0.70
ARR	freq %	537 17.12				61 3.89	9 0.57
VRQ	freq %	35 1.12					1 0.06
Total	freq	3 136					1 568

## The effect of PrP genotype on milk production and composition

The results of analysis of variance are presented in Table 2. The coefficients of determination for traits of milk quantity and quantities of milk ingredients range between 0.5793 and 0.6127 and those for milk composition traits between 0.0937 and 0.1629. Flock, parity and lactation length affected all studied traits significantly with the flock being the most important source of variance. The season of lambing also influenced all traits analysed with the exception of protein percentage. The year of lambing did not exhibit a significant effect on fat, lactose and dry matter percentage.

The PrP genotype was found to be a less important source of variance in the model. However, the significant effect of this source of variation was on milk yield (P=0.0190) and consequently on the yields of proteins and lactose produced in entire lactation. The yield of fat produced in lactation was not influenced by the genotype, neither were the concentrations of milk components with an exception of fat percentage.

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Trait	Genotype	Flock	Season	Parity	Lact. length	Year	R <sup>2</sup>
Milk, kg	0.0190*	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	0.6074
Proteins, kg	0.0339	< 0.0001	< 0.0001	< 0.0001	< 0.0001	0.0011	0.6127
Fat, kg	0.0965	< 0.0001	< 0.0001	0.0001	< 0.0001	< 0.0001	0.5821
Lactose, kg	0.0171	< 0.0001	< 0.0001	0.0001	< 0.0001	< 0.0001	0.5793
Proteins, %	0.1203	0.0001	0.8466	< 0.0001	< 0.0001	< 0.0001	0.1286
Fat, %	0.0098	< 0.0001	0.0032	0.0387	< 0.0001	0.1592	0.1629
Lactose, %	0.6940	< 0.0001	0.0136	< 0.0001	< 0.0001	0.7834	0.0946
Dry matter, %	0.1559	< 0.0001	0.0223	0.0003	< 0.0001	0.2119	0.0937

Table 2
Analysis of variance for milk yield, the yield of milk components in lactation and milk composition
Varianzanalyse der erfassten Milchmerkmale

Comparisons between genotypes revealed that the largest quantities of milk among homozygotes were produced by the ARR/ARR animals (Table 3), but the differences to other genotypes were not significant (P>0.05). The least productive were homozygotes ARH/ARH (210.5 kg in lactation); but because of the large standard error of that genotype, it was significantly different (P<0.05) only when compared to the most productive heterozygous animals AHQ/ARH (222.7 kg in lactation), which also produced significantly (P<0.05) more milk than ARQ/ARQ (210.5 kg) and ARR/ARQ (210.9 kg) genotypes. The quantities of proteins and lactose produced per lactation were directly dependent on milk quantity because, with the exception of fat content, the genotype had no influence on the concentration of milk ingredients. Differences among genotypes in protein and lactose yield show similar results as for milk yield. The percentage of milk fat was influenced by the PrP genotype, but the milk yield cancelled out that effect on fat yield and therefore PrP genotype had no significant effect on fat yield.

Comparisons between heterozygotes and their respective homozygotes (Table 4) showed that AHQ/ARH produced significantly (*P*<0.05) more milk (+14.19 kg per lactation), proteins (+0.75 kg per lactation) and lactose (+0.73) than the average of homozygotes ARH/ARH and AHQ/AHQ. Similarly, heterozygotes AHQ/ARQ (+6.94 kg milk, +0.38 kg fat and +0.36 kg lactose) and heterozygotes ARH/ARQ (+9.15 kg milk and +0.48 kg lactose) were superior (*P*<0.05) to their respective homozygotes AHQ/AHQ, ARQ/ARQ and ARH/ARH. However, the production of heterozygous genotypes ARR/ARH, ARR/ARH and ARR/ARQ was not statistically different from the average values of both homozygotes. In the total fat production and milk composition traits (protein, fat, lactose and dry matter percentage), heterozygous animals did not deviate from their respective homozygotes (Table 4) with the exception of heterozygote ARR/ARQ, which produced milk with significantly higher fat% (+0.119%) than ARR/ARR and ARQ/ARQ. However, the total fat produced per lactation by this heterozygote did not deviate from the average value of homozygotes, most likely because of its lower production of milk in lactation (–3.87).

### Discussion

The Bovška sheep is a Slovenian autochthonous dairy sheep breed with no recorded incidence of scrapie disease. As the risk for scrapie infection has increased in recent years,

<sup>\*</sup>numbers represent probabilities, R<sup>2</sup> coefficient of determination of the model

LSM  $\pm$  SE for quantities of milk yield and components and its percentages according to PrP genotypes LSM ± SE der Milchmengemerkmale und Anteile nach PrP Genotyp

2	Proteins, kg	Fat, kg	Lactose, kg	Proteins, %	Fat, %	Lactose, %	Dry matter, %
•	bc $11.53 \pm 0.22^{ab}$	$13.54 \pm 0.26$	$9.58 \pm 0.20^{ab}$	$5.45 \pm 0.04$	$6.48 \pm 0.06$ <sup>ac</sup>	$4.49 \pm 0.03$	$16.42 \pm 0.10$
_C.C ± 7.727 ± 3.5	$11.93 \pm 0.29^{a}$	$13.98 \pm 0.34$	$10.18 \pm 0.27^{ab}$	$5.39 \pm 0.05$	$6.37 \pm 0.08^{abcd}$	$4.54 \pm 0.03$	$16.30 \pm 0.13$
$^{\text{AHQ}/\text{ARQ}}$ 218.0 $\pm$ 2.2 $^{\text{ab}}$	b $11.67 \pm 0.12^a$	$13.85 \pm 0.14$	$9.91 \pm 0.11^{a}$	$5.40 \pm 0.02$	$6.45 \pm 0.03$ ac	$4.51 \pm 0.01$	16.36 v 0.05
RH/ARH 205.5 ± 6.3 <sup>b</sup>	$c$ 10.85 $\pm$ 0.34 <sup>b</sup>	$12.95 \pm 0.40$	$9.33 \pm 0.32^{ab}$	$5.30 \pm 0.06$	$6.43 \pm 0.10^{abcd}$	$4.53 \pm 0.04$	$16.27 \pm 0.16$
\text{ARH/ARQ} 217.2 ± 3.0 \text{\rightarrow}	$^{bc}$ 11.47 $\pm$ 0.16 $^{ab}$	$13.58 \pm 0.19$	$9.91 \pm 0.15^{a}$	$5.30 \pm 0.03$	$6.36 \pm 0.05^{\text{abd}}$	$4.53 \pm 0.02$	$16.20 \pm 0.07$
\rm RQ/ARQ \rm 210.5 ± 1.6°	$11.31 \pm 0.09^{b}$	$13.39 \pm 0.10$	$9.53 \pm 0.08^{b}$	$5.39 \pm 0.02$	$6.44 \pm 0.03^{\text{acd}}$	$4.49 \pm 0.01$	$16.33 \pm 0.04$
$RR/AHQ$ 211.7 $\pm$ 3.8 abc	$^{bc}$ 11.27 $\pm$ 0.21 $^{ab}$	$13.20 \pm 0.24$	$9.67 \pm 0.19^{ab}$	$5.35 \pm 0.04$	$6.35 \pm 0.06^{\text{abcd}}$	$4.53 \pm 0.02$	$16.23 \pm 0.09$
\text{\rm RR/ARH} 217.6 ± 4.9\alpha	$^{bc}$ 11.64 $\pm$ 0.26 $^{ab}$	$13.43 \pm 0.31$	$9.82 \pm 0.24^{ab}$	$5.36 \pm 0.05$	$6.21 \pm 0.08^{bd}$	$4.49 \pm 0.03$	$16.06 \pm 0.12$
$RR/ARQ$ 210.9 $\pm$ 2.1°	$11.27 \pm 0.11^{b}$	$13.47 \pm 0.13$	$9.57 \pm 0.10^{b}$	$5.37 \pm 0.02$	$6.49 \pm 0.03^{\circ}$	$4.51 \pm 0.01$	$16.36 \pm 0.05$
.RR/ARR 218.9 ± 4.4 abc	$^{bc}$ 11.65 $\pm$ 0.24 $^{ab}$	$13.55 \pm 0.28$	$9.93 \pm 0.22^{ab}$	$5.36 \pm 0.04$	$6.29 \pm 0.07^{d}$	$4.51 \pm 0.03$	$16.16 \pm 0.11$

Deviations of a heterozygote from their respective homozygotes (average value with its standard errors) Abweichungen der Heterozygoten von den entsprechenden Homozygoten (Mittelwert mit Standardfehler)

+:-:H			ceno	enotype		
Irait	AHQ/ARH	AHQ/ARQ	ARH/ARQ	ARR/AHQ	ARR/ARH	ARR/ARQ
Milk, kg	14.19 ±6.36*	$6.94 \pm 2.83*$	$9.15 \pm 4.29*$	$-3.51 \pm 4.66$	$5.41 \pm 6.10$	$-3.87 \pm 2.92$
Proteins, kg	$0.75 \pm 0.34^*$	$0.25 \pm 0.15*$	$0.39 \pm 0.23$	$-0.32 \pm 0.25$	$0.39 \pm 0.33$	$-0.21 \pm 0.16$
Fat, kg	$0.73 \pm 0.40$	$0.38 \pm 0.18$	$0.40 \pm 0.27$	$-0.34 \pm 0.29$	$0.18 \pm 0.39$	$0.00 \pm 0.18$
Lactose, kg	$0.73 \pm 0.32*$	$0.36 \pm 0.14^*$	$0.48 \pm 0.21^*$	$-0.09 \pm 0.23$	$0.19 \pm 0.30$	$-0.16 \pm 0.15$
Proteins, %	$0.012 \pm 0.063$	$-0.027 \pm 0.028$	$-0.043 \pm 0.042$	$-0.055 \pm 0.046$	$0.030 \pm 0.060$	$-0.006 \pm 0.029$
Fat, %	$-0.082 \pm 0.101$	$-0.012 \pm 0.045$	$-0.075 \pm 0.068$	$-0.038 \pm 0.073$	$-0.149 \pm 0.096$	$0.119 \pm 0.046$ *
Lactose, %	$0.032 \pm 0.041$	$0.022 \pm 0.018$	$0.024 \pm 0.028$	$0.030 \pm 0.030$	$-0.025 \pm 0.039$	$0.003 \pm 0.019$
Dry matter, %	$-0.046 \pm 0.156$	$-0.018 \pm 0.069$	$-0.102 \pm 0.105$	$-0.062 \pm 0.114$	$-0.153 \pm 0.149$	$0.167 \pm 0.071$

\*Deviation of heterozygote from the expected value differ significantly (P<0.05).

a programme for increasing genetic resistance was initiated in Europe. As such a selection programme can potentially have negative effects on certain production traits in particular breeds, we aimed here to determine the frequencies of PrP alleles and genotypes in Bovška sheep and evaluate their possible associations with milk production and composition traits.

The frequency of the most frequent allele ARQ (57.14%) is comparable with the frequencies found in other continental breeds (SALARIS *et al.* 2007, VAN KAAM *et al.* 2005, CASELLAS *et al.* 2006, WISNIEWSKA *et al.* 2006), although higher proportion of ARQ allele was found in the neighbouring Italian province Veneto (68.0%, DE MARCHI *et al.* 2005), in the Spanish Churra sheep (75.9%, ALVAREZ *et al.* 2006) and certain Northern German breeds (DRÖGEMÜLLER *et al.* 2001). The proportions of the desired ARR allele found in other continental breeds were mostly higher than in Bovška sheep (17.12%), from 18.2% in the Spanish Churra sheep (ALVAREZ *et al.* 2006) to 42.5% in the Sardinian sheep (SALARIS *et al.* 2007). However, the present study determined that the Bovška breed has among the lowest frequency of the susceptible VRQ allele (1.22%) reported, which can be viewed as an advantage for this breed. Higher frequencies of the VRQ allele were found in the Texel population (2 to 3%, BRANDSMA *et al.* 2004), in the Ripollesa breed (3%, CASELLAS *et al.* 2006) and in sheep from the Italian province Veneto (5.0%, DE MARCHI *et al.* 2005). However, these somewhat increased frequencies of more susceptible alleles are still much lower than those found in British breeds (RODEN *et al.* 2006).

Because of such a low proportion of the VRQ allele and a relatively low proportion of resistant ARR alleles (3.89% ARR/ARR homozygotes), Bovška sheep can be considered as an intermediately resistant breed to scrapie. The infection of Bovška sheep with scrapie could potentially threaten the breed and therefore increasing the ARR allele proportion could be beneficial.

The PrP genotype had a significant effect (*P*<0.05) on milk yield, protein yield and lactose yield per lactation. Yield of milk fat per lactation was not affected by the genotype, but the milk fat concentration was. However, PrP genotype had no significant (*P*>0.05) effect on the concentrations of proteins, lactose and total dry matter. Differences between the PrP genotypes in quantities of proteins and lactose were a consequence of differences in milk quantities produced per lactation and not the consequence of differences in concentrations of these two components. Furthermore, different yields of milk, produced by the different genotypes did not result in differences in total fat produced per lactation because of the effect of PrP genotype on fat concentration.

Hence the PrP genotype had a significant effect (P<0.05) on some productivity traits of Bovška sheep. The ARR/ARR were most productive among the studied homozygotes but the difference was statistically significant (P<0.05) only in comparison to ARH/ARH in milk and protein yield. Some heterozygotes showed better production results in milk yield, protein yield and lactose yield, compared to other genotypes. The most frequent homozygous genotype ARQ/ARQ produces less milk and proteins per lactation than some more productive heterozygotes, which implies a heterosis effect. For example, heterozygotes AHQ/ARH produced significantly more proteins (P<0.05) per lactation than homozygotes AHQ/AHQ and ARH/ARH, and heterozygotes AHQ/ARQ significantly (P<0.05) more milk fat than was expected from production of homozygotes AHQ/AHQ

and ARQ/ARQ. A similar pattern was found for lactose quantity per lactation, where homozygotes generally produced less lactose than heterozygotes.

The productivity of the most desired genotype ARR/ARR did not differ significantly (P>0.05) to other genotypes. It can therefore be expected that selection for resistant ARR alleles will not have an important direct negative effect on productivity of Bovška sheep population as has also been found in Sardinian sheep (SALARIS *et al.* 2007). The present study therefore suggests that moderate selection on genetic resistance to scrapie in Bovška sheep will most likely be beneficial without undesired consequences on milk production and composition traits. The productivity of the ARR homozygotes in this breed is at least at the same level as the productivity of the other genotypes. Also, the frequency of ARR is high enough to allow efficient simultaneous selection on genetic resistance against scrapie and for increased productivity. A relatively high frequency of the ARR allele also enables moderate selection pressure at the PrP locus that can, in turn, reduce potential negative effects of productivity due to reduced selection potential or inbreeding.

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